



Haemodialysis

CKJ REVIEW

Uraemic toxins and new methods to control their accumulation: game changers for the concept of dialysis adequacy

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Abstract

The current concept of an adequate dialysis based only on the dialysis process itself is rather limited. We now have considerable knowledge of uraemic toxicity and improved tools for limiting uraemic toxin accumulation. It is time to make use of these. A broader concept of adequacy that focusses on uraemic toxicity is required. As discussed in the present review, adequacy could be achieved by many different methods in combination with, or instead of, dialysis. These include preservation of renal function, dietary intake, reducing uraemic toxin generation rate and intestinal absorption, isolated ultrafiltration and extracorporeal adsorption of key uraemic toxins. A better measure of the quality of dialysis treatment would quantify the uraemic state in the patient using levels of a panel of key uraemic toxins. Treatment would focus on controlling uraemic toxicity while reducing harm or inconvenience to the patient. Delivering more dialysis might not be the best way to achieve this.

Key words: dialysis adequacy, new adequacy concepts, uraemic toxicity

Introduction

The current concept of dialysis adequacy focusses on urea and creatinine clearance (CrCl) by the dialysis system. This is rather simplistic and encourages providing more treatment by dialysis, ignoring other potentially fruitful strategies such as additional sessions of isolated ultrafiltration (UF), preserving renal function, reducing toxin generation rate, reducing toxin transfer from the gut, selectively adsorbing key toxins and modifying diet. These alternative methods could already be applied before dialysis starts or could be complementary to dialysis. According to the laws of mass action, biochemical effects of any toxin would be proportional to its concentration [1], which would depend as much on generation as on clearance. We now have considerable knowledge of uraemic toxicity and improved tools for limiting

their accumulation. It is time to make use of these. A broader concept of adequacy that focusses on uraemic toxicity is required.

Knowledge of uraemic toxicity has grown spectacularly over the past decades (Figure 1). Although barely discussed until late in the previous century, interest has increased exponentially since then. With the founding of the European Uraemic Toxin Workgroup (EUTox; www.uremic-toxins.org), an encyclopaedic list of uraemic retention solutes with their concentrations in uraemia became available [2]. A recent update confirmed the progressive increase in the number of identified retention solutes [3]. This can be attributed to improvements in analytic techniques and in the recent advances in the area of ‘-omics’, allowing profiling of the total proteome/metabolome within a biological sample [4, 5].

Received: February 21, 2015. Revised: April 21, 2015. Accepted: April 22, 2015

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Uraemic toxins are preferentially classified according to the physicochemical characteristics affecting their clearance during dialysis, which is still the main therapeutic option for their removal. Traditionally, this subdivision focusses on three types of molecules: the small water-soluble compounds [molecular weight (MW) < 500 Da], the larger ‘middle molecules’ (MW > 500 Da) and the protein-bound compounds [2]. Additionally, salt and water overload could be considered as causing uraemic toxicity. In the future, alternative classifications may be developed, based on new knowledge concerning, e.g. the generation of solutes as proposed in a recent review by Meijers *et al.* [6] pointing to new targets for decreasing levels of uraemic toxins (Figure 2). Ideally, nondialysis treatments to reduce uraemic toxicity could be started at earlier stages of chronic kidney disease (CKD).

Current adequacy methods

Current guidelines recommend quantifying dialysis by urea clearance. The evidence for this was based on the results of the

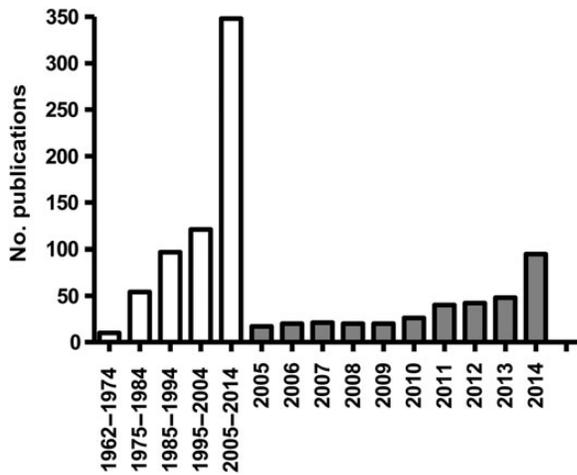


Fig. 1. Increasing number of publications on ‘uraemic toxins’ over the past decades.

national cooperative dialysis study (NCDS) [7], the first, and still one of the very few, randomized controlled trials (RCTs) designed to investigate the effect on outcome of varying dialysis dose. The NCDS randomized anuric patients dialysing thrice weekly into four groups according to target time-average blood urea nitrogen (BUN) and dialysis session length. The dialysis dose, quantified as the fractional volume cleared per dialysis (Kt/V), was prescribed for the individual patient to achieve the target BUN levels. Patients with higher urea generation rates were prescribed higher Kt/Vs to achieve their allocated BUN target. This study found a significantly reduced hospitalization rate ($P < 0.0001$) in the patients randomized to achieve low urea (time-averaged BUN 35 versus 75 mg/dL). Patients randomized to longer dialysis time (4.5 h) had ~50% reduced probability of being admitted compared with those treated by shorter dialysis (3.25 h), but this difference was not significant ($P = 0.06$). Subjects randomized to low BUN had to be given higher dialysis dose or have lower generation rate to achieve the BUN target. Since lower urea generation would have been due to lower dietary protein intake, usually associated with worse survival, the benefit of low BUN was likely to be due to the increased dose. The study concluded that achieving lower BUN levels was more effective at improving outcome than increasing session length. Secondary analysis of the NCDS, using a urea kinetic model to separate the effect of clearance and generation, suggested that the association between urea clearance as Kt/V and outcome was present at low clearance ($Kt/V < 0.9$) but was insignificant at clearance levels regarded as adequate by modern standards [8].

The Hemodialysis (HEMO) study is the only RCT designed to investigate the effect of higher dose of dialysis and outcome [9]. It found no benefit in increasing clearance above a Kt/V of 1.2, confirming the results of the NCDS. While there was no difference in outcome between the groups randomized to high dose versus standard dose, within each group, there was an association between poor outcome and failure to achieve the target Kt/V [10].

In peritoneal dialysis (PD), there is also no RCT evidence to support any specified Kt/V. The adequacy of PD in Mexico (ADEMEX) study showed no benefit of increasing Kt/V above 1.7 in anuric patients [11].

The decision when to start dialysis is a difficult and an important one for the patient. Ideally, we would have a measure

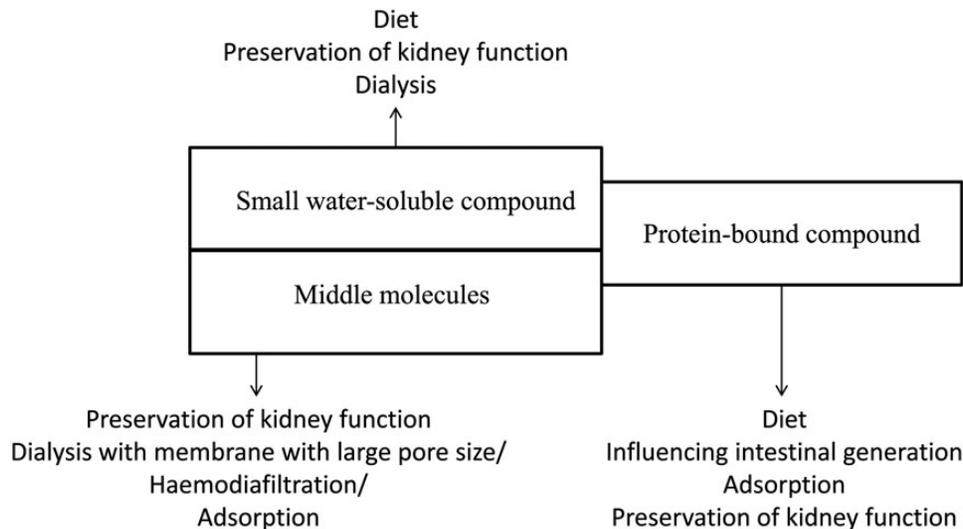


Fig. 2. Dialysis and nondialysis related techniques effective for controlling the levels of uraemic toxin.

of uraemic state so that we could start dialysis at the point when the advantages of dialysis outweigh the disadvantage. Estimation of glomerular filtration rate using serum creatinine (eGFR) has proven to be worse than useless as patients starting dialysis with low eGFR have a better outcome [12]. When eGFR is calculated from measurements of creatinine and urea clearance from urine collections, this association disappears [13]. It is possible that GFR measured by a more direct method would better predict outcome, but this has not yet been tested and would be difficult to apply in routine practice. Ideally, the uraemic state would be quantified by the direct measurements in plasma of one or more key uraemic toxins.

The initiation dialysis early and late (IDEAL) study prospectively investigated outcome in patients randomized to starting dialysis at a CrCl (estimated from serum creatinine) of 12 mL/min/1.73 m², whether or not the patients had symptoms of uraemia. The control group were patients starting dialysis when symptomatic or when CrCl dropped below 7. No difference was found [14]. It seems that survival is as good without dialysis but with even minimal levels of renal function compared with dialysis.

The evidence for controlling salt and water overload as a quantifiable and modifiable measure of the quality of dialysis seems at least as compelling as Kt/V [15]. Salt and water overload can be measured accurately by bioimpedance spectroscopy (BIS). Patients dialysing with longer dialysis sessions have improved outcome, compared with conventional treatment [16]. In a study comparing extended time and conventional dialysis, the worse outcome for short treatments was confined to patients who were salt and water overloaded [17]. This suggests that it is the better control of salt and water rather than the higher Kt/V, which is responsible for the improved outcome. Intensive control of fluid overload has been shown to reverse heart abnormalities.

Numerous studies have shown that residual renal function is associated with better outcome in dialysis patients. Where clearance measurements include the contribution of both dialysis and renal function, it is the contribution of the renal function, which has the dominant influence on outcome [18, 19].

So, it seems that, compared with normal renal function, a minimal dialysis providing <10% of the weekly urea clearance and much less than that for all other solutes will preserve life in the short term and avoid overt uraemic symptoms. But dialysis patients suffer from a range of long-term problems that reduce survival. Increasing the dose of dialysis, at least quantified by using standard methods, hardly improves outcome. With modern dialysis, it is easy to provide these minimal levels of clearance relatively noninvasively. Haemodialysis (HD) performed over 2 h three times weekly would deliver an adequate dose of dialysis with respect to small and middle molecule clearance, as defined in the NCDS study. However, with such short sessions, UF rates would be unacceptably high unless fluid weight gains between dialysis could be limited.

If we understood more about uraemic toxicity, we could use treatments other than dialysis to avoid or reduce toxicity. Where dialysis is required, we could fine-tune it to reduce the toxicity. This could improve the outcome for the patient or allow a less invasive and individualized treatment, specifically controlling the level of toxins causing problems for the patient, while limiting harm or inconvenience to the patient.

How to evaluate uraemic toxicity?

The evaluation of uraemic toxicity starts with identifying and quantifying the solutes that are present in uraemic biological fluids in abnormal concentrations. The biological effects of these potential uraemic toxins can be evaluated at relevant

concentrations in *in vitro/ex vivo* and/or *in vivo* experiments. In addition, clinical association studies can suggest a role of specific uraemic solutes in disease. The final approach is trying to decrease the concentrations *in vivo*, and only when an improvement of hard outcome of CKD patients is demonstrated, a causal relation is confirmed [20].

Analytical techniques

Individual uraemic retention solutes are analysed using colorimetric, fluorescence and high-performance liquid chromatographic (HPLC) methods. HPLC is also used to study groups of solutes sharing physical characteristics. As soon as '-omic' techniques, analysing total profiles of uraemic retentions solutes, became available, they were introduced into research on uraemic toxicity [4, 21–26]. In the context of uraemia, proteomics and metabolomics have been the main '-omic' applications [4, 21, 22, 24–28]. Proteomics is suited for the study of peptides and proteins (middle molecules) [29], while metabolomics focusses on small molecules. '-Omic' strategies are complementary and particularly useful as an approach for identifying pathways that are disturbed in a given pathology [30, 31].

Recently, proteomics have been applied in biomarker discovery, and a new proteome classifier assessing CKD and its prognosis has been proposed [32]. This study demonstrated that, although a high urinary protein excretion invariably resulted in renal failure progression, a low urinary protein excretion did not preclude death or dialysis. Even in patients without proteinuria, a low CKD273 score predicted renal failure progression within a follow-up period of 3.6 years [32]. This finding would need to be validated in independent cohorts before implementing into clinical practice [33].

Uraemic solutes identified in this way might not only be useful biomarkers but also real culprits in the progression of CKD and CKD-related cardiovascular disease (CVD).

When the concentrations of uraemic retention solutes applied in assays to evaluate their biological effects exceed those encountered in uraemia, conclusions on the solutes' toxicity might have relatively little clinical relevance [2]. Therefore, quantification of the confidently identified metabolites of interest should be performed by targeted methods before testing of the biological activity of uraemic retention solutes becomes possible (Table 1). Assessment of the pathophysiologic role of these newly detected metabolites will enable novel key culprits for the uraemic syndrome to be pointed out as the first step to pursue their specific removal.

Biological evaluation of toxicity of uraemic solutes

Small water-soluble compounds

Urea was the first uraemic retention solute to be identified and is amongst all uraemic retention solutes the one with the highest concentrations in the blood of uraemic patients. It reflects protein intake in the stable patient and has been used to assess nutrition and dialysis efficacy in renal patients. Toxicity of urea has remained elusive, and it has been thought that the uraemic syndrome was related to associated uraemic retention solutes but not to urea *per se*. However, more recently indirect toxic effect, via protein/albumin carbamylation [34] a risk factor for mortality in CKD [35], as well as limited direct toxic effects have been attributed to urea. Urea was found to induce the generation of reactive oxygen species (ROS) and insulin resistance *in vitro* and in mice [36]. In an *in vitro* study, Vaziri *et al.* showed that urea induced disruption of the intestinal epithelial barrier function by decreasing

Table 1. Key uraemic retention solutes

Uraemic retention solutes	MW (Da)	Normal concentration, mean (SD or range)	Uraemic concentration, mean (SD or range)	Ratio U/N
Small water-soluble				
Urea (g/L)	60	<0.4	2.3 (1.1)	5.7
ADMA (µg/L)	202	<60.6	878.7 (38.4)	14.5
SDMA (µg/L)	202	76.1 (21.0)	646.4 (606.0)	8.5
Middle molecules				
β2m (mg/L)	11 818	1.9 (1.6)	43.1 (18)	22.7
IL-6 (ng/L)	24 500	4.0	8.6 (3.7)	2.1
TNF-α (ng/L)	26 000	7.0	57.8 (10.8)	8.2
Protein-bound				
pCS (mg/L)	188	1.9 (1.3)	41 (13.3)	21.6
IS (mg/L)	212	0.53 (0.29)	44.5 (15.3)	84.0
IAA (mg/L)	175	0.5 (0.3)	2.4 (2.2)	4.8
HA (mg/L)	179	3.0 (2.0)	87.2 (61.7)	29.1
p-OHHA (mg/L)	195	NA	18.3 (6.6)	–

Extracted from [2, 3].

NA, not available; ADMA, asymmetric dimethylarginine; SDMA, symmetric dimethylarginine; β2m, beta 2 microglobulin; IL-6, interleukin-6; TNF-α, tumour necrosis factor-α; pCS, para-cresyl sulfate; IS, indoxyl sulfate; IAA, indole acetic acid; HA, hippuric acid; p-OHHA, para-hydroxyhippuric acid.

the expression of the tight junction proteins [Zona Occludens-1 (ZO-1), Claudin-1 and Occludin] [37]. Trecherel *et al.* explored regulatory proteins of apoptosis and showed an upregulation of Bcl2-associated death promoter (BAD), a pro-apoptotic protein [38].

Guanidines have been considered as uraemic toxins since the 1970s [39]. Guanidines are neurotoxins [40, 41]. They may also have cardiovascular toxicity since several guanidines are, based on leukocyte activation, pro-inflammatory at concentrations found in uraemia [42, 43]. Water-soluble guanidines are also responsible for the generation of other uraemic toxins like tumour necrosis factor-α (TNF-α) and interleukin-6 (IL-6), two middle molecules [42, 44]. The guanidines, asymmetric dimethylarginine (ADMA) and symmetric dimethylarginine (SDMA), are released from proteins that have been post-translationally methylated and subsequently hydrolysed. ADMA has for a long time been recognized as an inhibitor of nitric oxide synthase (NOS) causing endothelial dysfunction and vascular damage [45], a propensity that affects both the general and the uraemic population [46–48]. Infusion of ADMA in healthy volunteers, achieving a concentration as in uraemia, resulted in a decrease in cardiac output and a rise in vascular resistance [49]. SDMA, a structural analogue of ADMA, has long been considered inert [45, 50]. Its biologic activity was at first suggested by Bode-Boger *et al.* [51], showing a dose-dependent inhibition of NO synthesis mainly attributed to limiting the L-arginine supply to endothelial NOS. SDMA plays a role in leukocyte activation by enhancing generation of ROS, which is attributed to increased calcium influx via store-operated Ca²⁺ channels [52] and activation of nuclear factor (NF) κB resulting in cytokine production [44]. Inhibition of NF-κB activation by N-acetylcysteine (NAC) and ROS production with SKF96365 and captopril prevented leukocyte activation [44, 52]. Recently, Speer *et al.* [53] demonstrated that SDMA accumulates in high-density lipoprotein (HDL) particles from patients with CKD. This complex of HDL and SDMA is recognized by endothelial Toll-like receptor-2, leading to enhanced nicotinamide adenine dinucleotide phosphate-oxidase-dependent ROS production

and thereby reducing endothelial NO bioavailability *in vitro* and increasing arterial blood pressure *in vivo*. Hence, SDMA may be involved directly or indirectly in the pathogenesis of CVD via accumulation in HDL and seems neither to be inert nor to be a simple marker of renal function or CVD. However, the sole increase of SDMA by exogenous infusion in otherwise healthy mice affected neither renal function nor blood pressure or cardiac function [54].

Middle molecules

As mentioned earlier, the gradual increase of cytokines in CKD is, in addition to the reduced renal clearance, partly attributed to an increased generation in response to uraemic toxins [55, 56]. In clinical studies in CKD, pro-inflammatory cytokines are used as a hallmark of micro-inflammation [57]. The pathophysiological role of cytokines at concentrations as occurring in CKD is often neglected. It was recently demonstrated that, among several pro-inflammatory cytokines, TNF-α alone was pro-oxidative but only at high-range uraemic concentrations. The increase in ROS production could be blocked by adalimumab, although blocking had no effect on the oxidative stress in whole blood from HD patients, suggesting that other uraemic toxins than TNF-α are more crucial in this process [58].

Protein-bound compounds

Protein binding in CKD has been considered for some time, e.g. in the context of competition for drug binding [59]. It recently gained new interest as new dialysis techniques might have the potential to improve clearance of protein-bound toxins [60]. Protein-bound uraemic retention solutes have been studied extensively over the past decades with focus on their role in the increased susceptibility to infection and cardiovascular complications.

The biological effects of the prototype protein-bound solute, indoxyl sulfate (IS), have been studied the most. A recent systematic review [61] including 27 studies demonstrating pathophysiological effects of IS and/or p-cresyl sulfate (pCS) described their interference with several key metabolic processes involved in the uraemic syndrome. These included inflammation, oxidative stress, endothelial dysfunction, epithelial-to-mesenchymal transition, cardiac cell proliferation and renal tubular cell senescence. Since then, additional reports supporting the above evidence were published, covering increased crosstalk between leukocytes and endothelium, glycocalyx degradation and vascular leakage [62]; apoptosis of osteoblasts [63]; inhibition of drug metabolism [64]; induction of tubular endothelial growth factor receptor leading to tissue remodelling [65] and inhibition of breakdown of angiotensin II [66].

Similar effects were also described for other protein-bound toxins [67]. Indole acetic acid (IAA) was shown to inhibit endothelial progenitor cell production opposing their beneficial effect on vessel repair and neovascularization [68]. IAA induces endothelial inflammation and oxidative stress and activates an inflammatory AhR/p38MAPK/NF-κB pathway [69]. Recently, the ability of IAA to induce tissue factor production was associated with increased pro-coagulant activity [70, 71]. The induction of tissue factor occurred via the aryl hydrocarbon receptor pathway [71].

Recent metabolome studies repeatedly demonstrate increased levels of hippurates. Boelaert *et al.* demonstrated an increase, already from CKD Stage 3 on, of the known hippuric acid (HA) and 2-,3-,4-hydroxyhippuric acid. They also identified increased levels of an unknown aminohydroxyhippuric acid and of the sulphate and glucuronide conjugates of hydroxyhippuric acid [4]. HA was first isolated from horse urine, hence its name, and is a microbial co-metabolite. In general, literature on toxic effects of

hippurate is fairly old; somewhere along the way, interest in HA got lost. Satoh *et al.* demonstrated that subtotaly nephrectomized rats given HA in their drinking water showed a decrease in inulin clearance, pointing to glomerular dysfunction. This was supported by the significant increase in whole kidney sclerosis index. In addition, N-acetyl-glucosaminidase (NAG) excretion rate, an indicator of proximal tubular injury, was higher in the uraemic toxin-overloaded rats compared with the control rats [72]. More recently, HA was shown to inhibit the transport of two important efflux pumps expressed on human tubular cells [73]. Next to hippurate, hydroxyhippurates were increased in plasma from CKD patients. p-Hydroxyhippuric acid (p-OHHA) inhibits Ca^{2+} ATPases, needed for restoring intracellular Ca^{2+} homeostasis after cell activation. Increased intracellular Ca^{2+} modulates various polymorphonuclear leukocyte (PMNL) functions such as oxidative burst and degranulation as well as apoptosis as demonstrated by Cohen by the decrease in caspase activity in PMNL in the presence of p-OHHA [74].

Uraemic toxins and outcome

Several uraemic toxins have been linked to outcome in CKD patients or patients on dialysis [75].

ADMA concentration was correlated to intima media thickness, an index of vascular damage, in a dialysis population [46]. ADMA levels were found to be associated with high risk of death and cardiovascular events in predialysis patients [76–78] and dialysis patients [47]. A clinical study in 142 patients with different stages of CKD demonstrated a correlation of SDMA with TNF- α and IL-6 [44], which was markedly more significant than for ADMA [44]. Similarly, in a cohort of 288 dialysis patients, serum SDMA was a risk factor for death, in contrast to serum ADMA [79].

Elevated levels of cytokines and other inflammation markers have been related to all-cause and cardiovascular mortality in HD patients [80, 81]. In a population with advanced CKD, already having increased TNF- α concentrations, but not yet affected by possible negative effects of dialysis therapy, the concentration of TNF- α was not associated to adverse outcome [58], as was also shown for earlier stages of CKD [82]. This is in contrast to soluble TNF receptor 1 (sTNFR1) and sTNFR2, which are independently associated to all-cause mortality or an increased risk for cardiovascular events in advanced CKD irrespective of the cause of kidney disease [83], and IL-6, which has repeatedly been shown to be a strong predictor for outcome in CKD/dialysis [82, 84, 85].

For the protein-bound solutes, IS and pCS, highly significant associations between concentration and hard end points such as cardiovascular events, progression of renal failure and mortality have been demonstrated [86–91]. Serum IAA is an independent predictor of mortality and cardiovascular events in patients with CKD [69].

Finally, numerous studies have now linked the control of salt and water overload to outcome [15, 17, 92]. Intensive removal of excess fluid improves left ventricular hypertrophy. Most or all of the benefits of longer or more frequent dialysis sessions may be due to improved control of salt and water overload.

How to decrease concentrations/prevent accumulation of uraemic toxins?

Dietary modification

In anuric patients, fluid intake is usually driven by the need to dilute dietary salt. One litre is required for every 8 g of sodium chloride ingested [93]. Dietary sodium restriction would help

avoid salt and water overload and/or the need for UF. Similarly, restrictions in dietary potassium and phosphate are often recommended.

A very low-protein diet plus ketoacids (VLPD+) has been used to reduce urea generation and may delay or reduce the need for dialysis [94]. VLPD+ has also been shown to reduce the generation rate of IS, a known uraemic toxin [95].

Reducing absorption from the gut

Agents that bind phosphate or exchange phosphate for other solutes are used to limit phosphate accumulation in the majority of dialysis patients. Similarly, ion-exchange resins for potassium are occasionally used. Patiromer, an oral but nonadsorbed potassium binder, is effective in clinical trials [96].

Oral active charcoal, a nonspecific binder of organic toxins, is routinely used to treat poisoning. It has also been used successfully to control uraemia in patients who have refused dialysis [97] and to improve the abnormalities in gut barrier function in uraemia [98].

Recently, medicines have become available to limit absorption of specific classes of compound from the gut. These include orlistat for limiting fat absorption and lipoglyptin for limiting carbohydrate absorption. It is possible that, in the future, the limiting absorption of other toxins, more relevant to uraemia, will become available.

Reducing generation in the gut

A substantial part of the uraemic solutes is generated in the intestine as revealed by several studies, comparing the metabolome of germ-free mice versus mice with normal microbiota [99] and from HD patients with or without intact colon [100]. More recently, Holler *et al.* demonstrated the effect of prophylactic antibiotics on urinary IS in stem cell transplant recipients [101]. In spite of its importance, the intestinal microbiota is rarely taken into account in the context of uraemic toxicity and/or in the development/optimization of therapies. However, based on very few targeted studies, significant differences in the microbial composition in patients treated with HD [102] and PD [103] when compared with healthy controls have been reported. A recent untargeted study confirmed that uraemia alters the composition of the gut microbiome [104]. However, the effect of the altered microbial species composition on the metabolic activities linked to levels of protein-bound uraemic toxins in CKD is not known and needs further investigation, revealing whether the intestinal microbiota could be a possible future target even at earlier stages of CKD preventing generation rather than improving removal.

Preservation of kidney function

Even a severely damaged kidney may be capable of producing sufficient urine volume to prevent salt and water overload and avoid the need for UF. The urine volume may be increased, if required, by high-dose loop diuretics.

Residual renal function helps to control phosphate, beta2-microglobulin ($\beta_2\text{m}$) [105] and potassium [106]. In HD patients, the removal of protein-bound toxins may be entirely dependent on residual renal function. Survival is significantly associated with residual renal function in dialysis patients [19]. Multiple interventions can help preserve residual renal function. These include controlling blood sugar and blood pressure, avoiding nephrotoxic drugs and avoiding dehydration.

Influencing renal tubular handling of uraemic toxins may be another alternative and novel therapeutic approach to reduce their serum concentrations [107]. Transport of uraemic toxins

across the tubular cell membrane is facilitated by specific influx and efflux transporters. Changes in expression and/or function of influx transporters could decrease local toxicity to renal tubular cells [108, 109] and might also affect circulating concentrations if combined with effective efflux transport. Several uraemic toxins like indole-3-lactate, kynurenine and phenylsulfate are substrate to these transporters [99]. Drugs interfering with the function of these transporters, e.g. probenecid, inhibit the influx of uraemic toxins like IS, increasing viability of proximal tubular cells [110]. However, inhibition of these influx transporters will eventually contribute to further accumulation of uraemic toxins. In addition, expression of the organic anion transporters (OAT) 1, OAT3 and OAT polypeptide 4C1 (SLCO4C1) is shown to be decreased in CKD [111, 112]. Interestingly, Toyohara *et al.* demonstrated that the transcription of SLCO4C1 can be upregulated by statins, which leads to a higher expression on the cell membrane resulting in a decreased uraemic toxin concentration [112]. Mutsaers *et al.* recently reported that uraemic toxins inhibit substrate-specific uptake by both multidrug-resistance-associated protein (MRP4) and breast cancer-resistance protein (BCRP), two important renal efflux pumps [73]. This might again contribute to intracellular accumulation and toxicity. So, influx and efflux transporters might be an interesting target for trying to preserve tubular function, which is indispensable for the clearance of specifically protein-bound uraemic toxins.

Dialysis

Using knowledge of the principles of diffusion, clearance of any solute by any artificial dialysis system can be predicted [113, 114]. Existing dialysis systems, or their feasible enhancements, could be optimized to achieve target clearance for any uraemic toxin or group of toxin.

Low-molecular-weight toxins are easily cleared by HD. Levels of toxins similar to that found in patients with normal renal function could be achieved by daily 8-h sessions of high-efficiency dialysis.

Higher-molecular-weight toxins can also be removed effectively by dialysis, as long as they are not bound to protein and the molecules are small enough to pass through the dialyser membrane's pores. Membranes that have a pore radius just smaller than that of albumin are available. Due to the lower rate of diffusion of these larger toxins, efficient clearance rates require larger membrane surface area and are helped by convection or fluid flow across the membrane. Haemodiafiltration, in which up to 100 mL/min of plasma water is filtered across the membrane, could reduce the levels of larger solutes to close to normal levels with daily 8-h treatments.

For protein-bound toxins, only the unbound fraction can be removed by HD or filtration. Clearance of bound toxin requires removal and replacement of the plasma-binding protein (usually albumin), using a membrane that is porous to albumin [115]. The plasma protein can be stripped of the bound toxin by contact with a competitive binding agent, before re-infusion of the plasma proteins into the patient. Systems capable of removing bound toxin are currently available but expensive. Current dialyser membranes bind certain toxins (e.g. β_2m). Dialysers could be modified to include a matrix that would adsorb specific uraemic toxins. Since the matrix would be in direct contact with plasma proteins, these could adsorb bound toxin. A carbon-based matrix has been shown to reduce the levels of protein-bound toxins IS and pCS *in vitro* [116, 117].

Excess salt and water overload can be removed by UF. Rapid UF causes ischaemia by increasing blood viscosity and reducing

blood pressure [118]. Equipment to remove fluid by UF without dialysis is much simpler, cheaper and portable compared with dialysis. Isolated UF can be powered by the patient's arterial blood pressure and needs no water or chemical supplies. Salt and water overload could be more easily avoided by more frequent or continuous UF. Harmful effects of rapid UF can be avoided by longer or continuous UF. Longer or more frequent treatments may be more acceptable to the patient using a portable or even implantable UF device [119].

Potential new adequacy concepts

Kt/V is useful to calibrate the dialysis process, to verify that a dialysis has been delivered as prescribed and as a measure of dialysis dose, but the achievement of a universally specified Kt/V value should not be an objective in itself. Kt/V does not predict levels of any uraemic toxin [120]. It does not even predict levels of urea. We need ways to quantify the uraemic state, so we can abandon Kt/V as a measure of dialysis adequacy.

Since toxicity should be proportional to concentration of the toxin, the quality of dialysis would be assessed on the concentrations of toxins in the patient. Adequately low levels could be achieved by limiting its generation, preserving or enhancing renal clearance as well as or instead of dialysis. Excess salt and water would be considered as a key 'toxin', and an adequate dialysis would limit this without excessive UF rate.

Concentrations of toxins may be predicted using knowledge of the toxin's generation rate and clearance. Manufacturers of dialysers would provide sufficient data to allow clearance of key toxins to be predicted.

To some extent, this concept of adequacy has already been implemented for PD, where renal clearance and ability to control fluid overload are known to be crucial and small solute clearance by dialysis relatively unimportant.

Conclusion

The current concept of an adequate dialysis based only on the dialysis process itself is rather limited. It would be better to include factors within the patient such as dietary intake, generation and renal function. Adequacy could be achieved by many different methods in combination with, or instead of, dialysis. These include preservation of renal function, isolated UF, extracorporeal adsorption of key toxins, modifying diet, reducing intestinal absorption and toxin generation rate.

A better measure of the quality of end-stage renal disease treatment would quantify the uraemic state in the patient using levels of a panel of key toxins. Treatment would focus on controlling uraemic toxicity while reducing harm or inconvenience to the patient. Delivering more dialysis might not be the best way to achieve this.

Conflict of interest statement

None declared.

References

1. Guldberg CM, Waage P. *Studies Concerning Affinity*. Forhandlinger: Videnskabs-Selskabet i Christiania 35, 1864
2. Vanholder R, De Smet R, Glorieux G *et al.* Review on uremic toxins: classification, concentration, and interindividual variability. *Kidney Int* 2003; 63: 1934–1943

- Duranton F, Cohen G, De Smet R et al. Normal and pathologic concentrations of uremic toxins. *J Am Soc Nephrol* 2012; 23: 1258–1270
- Boelaert J, t'Kindt R, Schepers E et al. State-of-the-art non-targeted metabolomics in the study of chronic kidney disease. *Metabolomics* 2013; 10: 425–442
- Weissinger EM, Kaiser T, Meert N et al. Proteomics: a novel tool to unravel the patho-physiology of uraemia. *Nephrol Dial Transplant* 2004; 19: 3068–3077
- Meijers B, Glorieux G, Poesen R et al. Nonextracorporeal methods for decreasing uremic solute concentration: a future way to go? *Semin Nephrol* 2014; 34: 228–243
- Lowrie EG, Laird NM, Parker TF et al. Effect of the hemodialysis prescription of patient morbidity: report from the national cooperative dialysis study. *N Engl J Med* 1981; 305: 1176–1181
- Gotch FA, Sargent JA. A mechanistic analysis of the national cooperative dialysis study (NCDS). *Kidney Int* 1985; 28: 526–534
- Eknoyan G, Beck GJ, Cheung AK et al. Effect of dialysis dose and membrane flux in maintenance hemodialysis. *N Engl J Med* 2002; 347: 2010–2019
- Greene T, Daugirdas J, Depner T et al. Association of achieved dialysis dose with mortality in the hemodialysis study: an example of “dose-targeting bias”. *J Am Soc Nephrol* 2005; 16: 3371–3380
- Paniagua R, Amato D, Vonesh E et al. Effects of increased peritoneal clearances on mortality rates in peritoneal dialysis: ADEMEX, a prospective, randomized, controlled trial. *J Am Soc Nephrol* 2002; 13: 1307–1320
- Fink JC, Burdick RA, Kurth SJ et al. Significance of serum creatinine values in new end-stage renal disease patients. *Am J Kidney Dis* 1999; 34: 694–701
- Grootendorst DC, Michels WM, Richardson JD et al. The MDRD formula does not reflect GFR in ESRD patients. *Nephrol Dial Transplant* 2011; 26: 1932–1937
- Cooper BA, Branley P, Bulfone L et al. A randomized, controlled trial of early versus late initiation of dialysis. *N Engl J Med* 2010; 363: 609–619
- Tsai YC, Chiu YW, Tsai JC et al. Association of fluid overload with cardiovascular morbidity and all-cause mortality in stages 4 and 5 CKD. *Clin J Am Soc Nephrol* 2015; 10: 39–46
- Tentori F, Zhang J, Li Y et al. Longer dialysis session length is associated with better intermediate outcomes and survival among patients on in-center three times per week hemodialysis: results from the dialysis outcomes and practice patterns study (DOPPS). *Nephrol Dial Transplant* 2012; 27: 4180–4188
- Chazot C, Wabel P, Chamney P et al. Importance of normohydration for the long-term survival of haemodialysis patients. *Nephrol Dial Transplant* 2012; 27: 2404–2410
- Bargman JM, Thorpe KE, Churchill DN. Relative contribution of residual renal function and peritoneal clearance to adequacy of dialysis: a reanalysis of the CANUSA study. *J Am Soc Nephrol* 2001; 12: 2158–2162
- Termorshuizen F, Dekker FW, van Manen JG et al. Relative contribution of residual renal function and different measures of adequacy to survival in hemodialysis patients: an analysis of the Netherlands cooperative study on the adequacy of dialysis (NECOSAD)-2. *J Am Soc Nephrol* 2004; 15: 1061–1070
- Glorieux G, Vanholder R. New uremic toxins—which solutes should be removed? *Contrib Nephrol* 2011; 168: 117–128
- Mutsaers HA, Engelke UF, Wilmer MJ et al. Optimized metabolomic approach to identify uremic solutes in plasma of stage 3–4 chronic kidney disease patients. *PLoS One* 2013; 8: e71199
- Naseeb U, Shafqat J, Jagerbrink T et al. Proteome patterns in uremic plasma. *Blood Purif* 2008; 26: 561–568
- Perco P, Muhlberger I, Mayer G et al. Linking transcriptomic and proteomic data on the level of protein interaction networks. *Electrophoresis* 2010; 31: 1780–1789
- Schiffer E, Mischak H, Vanholder RC. Exploring the uremic toxins using proteomic technologies. *Contrib Nephrol* 2008; 160: 159–171
- Shah VO, Townsend RR, Feldman HI et al. Plasma metabolomic profiles in different stages of CKD. *Clin J Am Soc Nephrol* 2013; 8: 363–370
- Toyohara T, Akiyama Y, Suzuki T et al. Metabolomic profiling of uremic solutes in CKD patients. *Hypertens Res* 2010; 33: 944–952
- Glorieux G, Helling R, Henle T et al. In vitro evidence for immune activating effect of specific AGE structures retained in uremia. *Kidney Int* 2004; 66: 1873–1880
- Mullen W, Saigusa D, Abe T et al. Proteomics and metabolomics as tools to unravel novel culprits and mechanisms of uremic toxicity: instrument or hype? *Semin Nephrol* 2014; 34: 180–190
- Rodriguez-Suarez E, Siwy J, Zurbig P et al. Urine as a source for clinical proteome analysis: from discovery to clinical application. *Biochim Biophys Acta* 2014; 1844: 884–898
- Dunn WB, Bailey NJ, Johnson HE. Measuring the metabolome: current analytical technologies. *Analyst* 2005; 130: 606–625
- Goodacre R, Vaidyanathan S, Dunn WB et al. Metabolomics by numbers: acquiring and understanding global metabolite data. *Trends Biotechnol* 2004; 22: 245–252
- Argiles A, Siwy J, Duranton F et al. CKD273, a new proteomics classifier assessing CKD and its prognosis. *PLoS One* 2013; 8: e62837
- Mischak H, Ioannidis JP, Argiles A et al. Implementation of proteomic biomarkers: making it work. *Eur J Clin Invest* 2012; 42: 1027–1036
- Kraus LM, Kraus AP Jr. Carbamoylation of amino acids and proteins in uremia. *Kidney Int Suppl* 2001; 78: S102–S107
- Berg AH, Drechsler C, Wenger J et al. Carbamylation of serum albumin as a risk factor for mortality in patients with kidney failure. *Sci Transl Med* 2013; 5: 175ra29
- D'Apolito M, Du X, Zong H et al. Urea-induced ROS generation causes insulin resistance in mice with chronic renal failure. *J Clin Invest* 2010; 120: 203–213
- Vaziri ND, Yuan J, Norris K. Role of urea in intestinal barrier dysfunction and disruption of epithelial tight junction in chronic kidney disease. *Am J Nephrol* 2013; 37: 1–6
- Trecherel E, Godin C, Louandre C et al. Upregulation of BAD, a pro-apoptotic protein of the BCL2 family, in vascular smooth muscle cells exposed to uremic conditions. *Biochem Biophys Res Commun* 2012; 417: 479–483
- Carroll HJ. Energy production and utilization by human platelets in the presence of some guanidines and phenols (uremic toxins) that inhibit aggregation. *Thromb Diath Haemorrh* 1975; 34: 63–71
- D'Hooge R, Pei YQ, Marescau B et al. Convulsive action and toxicity of uremic guanidino compounds: behavioral assessment and relation to brain concentration in adult mice. *J Neurol Sci* 1992; 112: 96–105
- D'Hooge R, Van de Vijver G, Van Bogaert PP et al. Involvement of voltage- and ligand-gated Ca²⁺ channels in the

- neuroexcitatory and synergistic effects of putative uremic neurotoxins. *Kidney Int* 2003; 63: 1764–1775
42. Glorieux GL, Dhondt AW, Jacobs P et al. In vitro study of the potential role of guanidines in leukocyte functions related to atherogenesis and infection. *Kidney Int* 2004; 65: 2184–2192
 43. Schepers E, Glorieux G, Dou L et al. Guanidino compounds as cause of cardiovascular damage in chronic kidney disease: an in vitro evaluation. *Blood Purif* 2010; 30: 277–287
 44. Schepers E, Barreto DV, Liabeuf S et al. Symmetric dimethylarginine as a proinflammatory agent in chronic kidney disease. *Clin J Am Soc Nephrol* 2011; 6: 2374–2383
 45. Leiper J, Vallance P. Biological significance of endogenous methylarginines that inhibit nitric oxide synthases. *Cardiovasc Res* 1999; 43: 542–548
 46. Zoccali C, Benedetto FA, Maas R et al. Asymmetric dimethylarginine, C-reactive protein, and carotid intima-media thickness in end-stage renal disease. *J Am Soc Nephrol* 2002; 13: 490–496
 47. Zoccali C, Bode-Boger S, Mallamaci F et al. Plasma concentration of asymmetrical dimethylarginine and mortality in patients with end-stage renal disease: a prospective study. *Lancet* 2001; 358: 2113–2117
 48. Meinitzer A, Seelhorst U, Wellnitz B et al. Asymmetrical dimethylarginine independently predicts total and cardiovascular mortality in individuals with angiographic coronary artery disease (the Ludwigshafen risk and cardiovascular health study). *Clin Chem* 2007; 53: 273–283
 49. Kielstein JT, Impraim B, Simmel S et al. Cardiovascular effects of systemic nitric oxide synthase inhibition with asymmetrical dimethylarginine in humans. *Circulation* 2004; 109: 172–177
 50. Vallance P, Leone A, Calver A et al. Accumulation of an endogenous inhibitor of nitric oxide synthesis in chronic renal failure. *Lancet* 1992; 339: 572–575
 51. Bode-Boger SM, Scalera F, Kielstein JT et al. Symmetrical dimethylarginine: a new combined parameter for renal function and extent of coronary artery disease. *J Am Soc Nephrol* 2006; 17: 1128–1134
 52. Schepers E, Glorieux G, Dhondt A et al. Role of symmetric dimethylarginine in vascular damage by increasing ROS via store-operated calcium influx in monocytes. *Nephrol Dial Transplant* 2009; 24: 1429–1435
 53. Speer T, Rohrer L, Blyszczuk P et al. Abnormal high-density lipoprotein induces endothelial dysfunction via activation of Toll-like receptor-2. *Immunity* 2013; 38: 754–768
 54. Veldink H, Faulhaber-Walter R, Park JK et al. Effects of chronic SDMA infusion on glomerular filtration rate, blood pressure, myocardial function and renal histology in C57BL6/J mice. *Nephrol Dial Transplant* 2013; 28: 1434–1439
 55. Bemelmans MH, Gouma DJ, Buurman WA. Influence of nephrectomy on tumor necrosis factor clearance in a murine model. *J Immunol* 1993; 150: 2007–2017
 56. Garibotto G, Sofia A, Balbi M et al. Kidney and splanchnic handling of interleukin-6 in humans. *Cytokine* 2007; 37: 51–54
 57. Carrero JJ, Park SH, Axelsson J et al. Cytokines, atherogenesis, and hypercatabolism in chronic kidney disease: a dreadful triad. *Semin Dial* 2009; 22: 381–386
 58. Neiryneck N, Glorieux G, Schepers E et al. Pro-inflammatory cytokines and leukocyte oxidative burst in chronic kidney disease: culprits or innocent bystanders? *Nephrol Dial Transplant* 2015; 30: 943–951
 59. Vanholder R, Van LN, De SR et al. Drug protein binding in chronic renal failure: evaluation of nine drugs. *Kidney Int* 1988; 33: 996–1004
 60. Bohringer F, Jankowski V, Gajjala PR et al. Release of uremic retention solutes from protein binding by hypertonic predilution hemodiafiltration. *ASAIO J* 2015; 61: 55–60
 61. Vanholder R, Schepers E, Pletinck A et al. The uremic toxicity of indoxyl sulfate and p-cresyl sulfate: a systematic review. *J Am Soc Nephrol* 2014; 25: 1897–1907
 62. Pletinck A, Glorieux G, Schepers E et al. Protein-bound uremic toxins stimulate crosstalk between leukocytes and vessel wall. *J Am Soc Nephrol* 2013; 24: 1981–1994
 63. Kim YH, Kwak KA, Gil HW et al. Indoxyl sulfate promotes apoptosis in cultured osteoblast cells. *BMC Pharmacol Toxicol* 2013; 14: 60
 64. Barnes KJ, Rowland A, Polasek TM et al. Inhibition of human drug-metabolising cytochrome P450 and UDP-glucuronosyltransferase enzyme activities in vitro by uremic toxins. *Eur J Clin Pharmacol* 2014; 70: 1097–1106
 65. Sun CY, Young GH, Hsieh YT et al. Protein-bound uremic toxins induce tissue remodeling by targeting the EGF receptor. *J Am Soc Nephrol* 2014; 26: 281–290
 66. Ng HY, Yisireyili M, Saito S et al. Indoxyl sulfate downregulates expression of Mas receptor via OAT3/AhR/Stat3 pathway in proximal tubular cells. *PLoS One* 2014; 9: e91517
 67. Sirich TL, Meyer TW, Gondouin B et al. Protein-bound molecules: a large family with a bad character. *Semin Nephrol* 2014; 34: 106–117
 68. Jourde-Chiche N, Dou L, Sabatier F et al. Levels of circulating endothelial progenitor cells are related to uremic toxins and vascular injury in hemodialysis patients. *J Thromb Haemost* 2009; 7: 1576–1584
 69. Dou L, Sallee M, Cerini C et al. The cardiovascular effect of the uremic solute indole-3 acetic acid. *J Am Soc Nephrol* 2015; 26: 876–887
 70. Chitalia VC, Shivanna S, Martorell J et al. Uremic serum and solutes increase post-vascular interventional thrombotic risk through altered stability of smooth muscle cell tissue factor. *Circulation* 2013; 127: 365–376
 71. Gondouin B, Cerini C, Dou L et al. Indolic uremic solutes increase tissue factor production in endothelial cells by the aryl hydrocarbon receptor pathway. *Kidney Int* 2013; 84: 733–744
 72. Satoh M, Hayashi H, Watanabe M et al. Uremic toxins overload accelerates renal damage in a rat model of chronic renal failure. *Nephron Exp Nephrol* 2003; 95: e111–e118
 73. Mutsaers HA, van den Heuvel LP, Ringens LH et al. Uremic toxins inhibit transport by breast cancer resistance protein and multidrug resistance protein 4 at clinically relevant concentrations. *PLoS One* 2011; 6: e18438
 74. Cohen G, Raupachova J, Wimmer T et al. The uraemic retention solute para-hydroxy-hippuric acid attenuates apoptosis of polymorphonuclear leukocytes from healthy subjects but not from haemodialysis patients. *Nephrol Dial Transplant* 2008; 23: 2512–2519
 75. Liabeuf S, Neiryneck N, Drueke TB et al. Clinical studies and chronic kidney disease: what did we learn recently? *Semin Nephrol* 2014; 34: 164–179
 76. Fliser D, Kronenberg F, Kielstein JT et al. Asymmetric dimethylarginine and progression of chronic kidney disease: the mild to moderate kidney disease study. *J Am Soc Nephrol* 2005; 16: 2456–2461
 77. Ravani P, Tripepi G, Malberti F et al. Asymmetrical dimethylarginine predicts progression to dialysis and death in patients with chronic kidney disease: a competing risks modeling approach. *J Am Soc Nephrol* 2005; 16: 2449–2455
 78. Shi B, Ni Z, Zhou W et al. Circulating levels of asymmetric dimethylarginine are an independent risk factor for left

- ventricular hypertrophy and predict cardiovascular events in pre-dialysis patients with chronic kidney disease. *Eur J Intern Med* 2010; 21: 444–448
79. Aucella F, Maas R, Vigilante M *et al.* Methylarginines and mortality in patients with end stage renal disease: a prospective cohort study. *Atherosclerosis* 2009; 207: 541–545
 80. Qureshi AR, Alvestrand A, vino-Filho JC *et al.* Inflammation, malnutrition, and cardiac disease as predictors of mortality in hemodialysis patients. *J Am Soc Nephrol* 2002; 13(Suppl 1): S28–S36
 81. Zimmermann J, Herrlinger S, Pruy A *et al.* Inflammation enhances cardiovascular risk and mortality in hemodialysis patients. *Kidney Int* 1999; 55: 648–658
 82. Barreto DV, Barreto FC, Liabeuf S *et al.* Plasma interleukin-6 is independently associated with mortality in both hemodialysis and pre-dialysis patients with chronic kidney disease. *Kidney Int* 2010; 77: 550–556
 83. Neiryck N, Glorieux G, Schepers E *et al.* Soluble tumor necrosis factor receptor 1 and 2 predict outcomes in advanced chronic kidney disease: a prospective cohort study. *PLoS One* 2015; 10: e0122073
 84. Pecoits-Filho R, Barany P, Lindholm B *et al.* Interleukin-6 is an independent predictor of mortality in patients starting dialysis treatment. *Nephrol Dial Transplant* 2002; 17: 1684–1688
 85. Panichi V, Rizza GM, Paoletti S *et al.* Chronic inflammation and mortality in haemodialysis: effect of different renal replacement therapies. Results from the RISCAVID study. *Nephrol Dial Transplant* 2008; 23: 2337–2343
 86. Barreto FC, Barreto DV, Liabeuf S *et al.* Serum indoxyl sulfate is associated with vascular disease and mortality in chronic kidney disease patients. *Clin J Am Soc Nephrol* 2009; 4: 1551–1558
 87. Chiu CA, Lu LF, Yu TH *et al.* Increased levels of total p-cresylsulphate and indoxyl sulphate are associated with coronary artery disease in patients with diabetic nephropathy. *Rev Diabet Stud* 2010; 7: 275–284
 88. Liabeuf S, Barreto DV, Barreto FC *et al.* Free p-cresylsulphate is a predictor of mortality in patients at different stages of chronic kidney disease. *Nephrol Dial Transplant* 2010; 25: 1183–1191
 89. Wang CP, Lu LF, Yu TH *et al.* Serum levels of total p-cresylsulphate are associated with angiographic coronary atherosclerosis severity in stable angina patients with early stage of renal failure. *Atherosclerosis* 2010; 211: 579–583
 90. Wu IW, Hsu KH, Lee CC *et al.* p-Cresyl sulphate and indoxyl sulphate predict progression of chronic kidney disease. *Nephrol Dial Transplant* 2011; 26: 938–947
 91. Wu IW, Hsu KH, Hsu HJ *et al.* Serum free p-cresyl sulfate levels predict cardiovascular and all-cause mortality in elderly hemodialysis patients—a prospective cohort study. *Nephrol Dial Transplant* 2012; 27: 1169–1175
 92. Velasco N, Chamney P, Wabel P *et al.* Optimal fluid control can normalize cardiovascular risk markers and limit left ventricular hypertrophy in thrice weekly dialysis patients. *Hemodial Int* 2012; 16: 465–472
 93. Lindley EJ. Reducing sodium intake in hemodialysis patients. *Semin Dial* 2009; 22: 260–263
 94. Caria S, Cupisti A, Sau G *et al.* The incremental treatment of ESRD: a low-protein diet combined with weekly hemodialysis may be beneficial for selected patients. *BMC Nephrol* 2014; 15: 172
 95. Marzocco S, Dal PF, Di ML *et al.* Very low protein diet reduces indoxyl sulfate levels in chronic kidney disease. *Blood Purif* 2013; 35: 196–201
 96. Weir MR, Bakris GL, Bushinsky DA *et al.* Patiromer in patients with kidney disease and hyperkalemia receiving RAAS inhibitors. *N Engl J Med* 2015; 372: 211–221
 97. Musso CG, Michelangelo H, Reynaldi J *et al.* Combination of oral activated charcoal plus low protein diet as a new alternative for handling in the old end-stage renal disease patients. *Saudi J Kidney Dis Transpl* 2010; 21: 102–104
 98. Vaziri ND, Yuan J, Khazaeli M *et al.* Oral activated charcoal adsorbent (AST-120) ameliorates chronic kidney disease-induced intestinal epithelial barrier disruption. *Am J Nephrol* 2013; 37: 518–525
 99. Wikoff WR, Nagle MA, Kouznetsova VL *et al.* Untargeted metabolomics identifies enterobiome metabolites and putative uremic toxins as substrates of organic anion transporter 1 (Oat1). *J Proteome Res* 2011; 10: 2842–2851
 100. Aronov PA, Luo FJ, Plummer NS *et al.* Colonic contribution to uremic solutes. *J Am Soc Nephrol* 2011; 22: 1769–1776
 101. Holler E, Butzhammer P, Schmid K *et al.* Metagenomic analysis of the stool microbiome in patients receiving allogeneic stem cell transplantation: loss of diversity is associated with use of systemic antibiotics and more pronounced in gastrointestinal graft-versus-host disease. *Biol Blood Marrow Transplant* 2014; 20: 640–645
 102. Hida M, Aiba Y, Sawamura S *et al.* Inhibition of the accumulation of uremic toxins in the blood and their precursors in the feces after oral administration of Lebenin, a lactic acid bacteria preparation, to uremic patients undergoing hemodialysis. *Nephron* 1996; 74: 349–355
 103. Wang IK, Lai HC, Yu CJ *et al.* Real-time PCR analysis of the intestinal microbiotas in peritoneal dialysis patients. *Appl Environ Microbiol* 2012; 78: 1107–1112
 104. Vaziri ND, Wong J, Pahl M *et al.* Chronic kidney disease alters intestinal microbial flora. *Kidney Int* 2013; 83: 308–315
 105. Penne EL, van der Weerd NC, Blankestijn PJ *et al.* Role of residual kidney function and convective volume on change in beta2-microglobulin levels in hemodiafiltration patients. *Clin J Am Soc Nephrol* 2010; 5: 80–86
 106. Vilar E, Farrington K. Emerging importance of residual renal function in end-stage renal failure. *Semin Dial* 2011; 24: 487–494
 107. Masereeuw R, Mutsaers HA, Toyohara T *et al.* The kidney and uremic toxin removal: glomerulus or tubule? *Semin Nephrol* 2014; 34: 191–208
 108. Poveda J, Sanchez-Nino MD, Glorieux G *et al.* p-Cresyl sulphate has pro-inflammatory and cytotoxic actions on human proximal tubular epithelial cells. *Nephrol Dial Transplant* 2014; 29: 56–64
 109. Watanabe H, Miyamoto Y, Honda D *et al.* p-Cresyl sulfate causes renal tubular cell damage by inducing oxidative stress by activation of NADPH oxidase. *Kidney Int* 2013; 83: 582–592
 110. Enomoto A, Takeda M, Tojo A *et al.* Role of organic anion transporters in the tubular transport of indoxyl sulfate and the induction of its nephrotoxicity. *J Am Soc Nephrol* 2002; 13: 1711–1720
 111. Deguchi T, Takemoto M, Uehara N *et al.* Renal clearance of endogenous hippurate correlates with expression levels of renal organic anion transporters in uremic rats. *J Pharmacol Exp Ther* 2005; 314: 932–938
 112. Toyohara T, Suzuki T, Morimoto R *et al.* SLCO4C1 transporter eliminates uremic toxins and attenuates hypertension and renal inflammation. *J Am Soc Nephrol* 2009; 20: 2546–2555
 113. Einstein A. Über die von der molekularkinetischen Theorie der Wärme geforderte Bewegung von in ruhenden Flüssigkeiten suspendierten Teilchen. *Annalen der Physik* 1905; 17: 549–560

114. Werynski A, Waniewski J. Theoretical description of mass transport in medical membrane devices. *Artif Organs* 1995; 19: 420–427
115. Patzer JF. Thermodynamic considerations in solid adsorption of bound solutes for patient support in liver failure. *Artif Organs* 2008; 32: 499–508
116. Sandeman SR, Howell CA, Phillips GJ et al. An adsorbent monolith device to augment the removal of uraemic toxins during haemodialysis. *J Mater Sci Mater Med* 2014; 25: 1589–1597
117. Sarnatskaya VV, Yushko LA, Sakhno LA et al. New approaches to the removal of protein-bound toxins from blood plasma of uremic patients. *Artif Cells Blood Substit Immobil Biotechnol* 2007; 35: 287–308
118. Burton JO, Jefferies HJ, Selby NM et al. Hemodialysis-induced cardiac injury: determinants and associated outcomes. *Clin J Am Soc Nephrol* 2009; 4: 914–920
119. Gura V, Ronco C, Nalesso F et al. A wearable hemofilter for continuous ambulatory ultrafiltration. *Kidney Int* 2008; 73: 497–502
120. Eloit S, Van Biesen W, Glorieux G et al. Does the adequacy parameter Kt/V_{urea} reflect uremic toxin concentrations in hemodialysis patients? *PLoS ONE* 2013; 8: e76838